

Human Brain Microvascular Endothelial Cells (HBMECs)

Catalog #: NB-26-00185-500

Cell #: > 5X10⁵ cells

Storage: 37°C CO₂ incubator or Liquid Nitrogen

Product Format: Frozen

General Information

HBMECs were isolated from normal human brain tissue. Passage 1, cells are ship on dry ice in a vial. ENDO-Growth Media is recommended for culture. Cells have an average additional population doubling levels >15 when cultured.

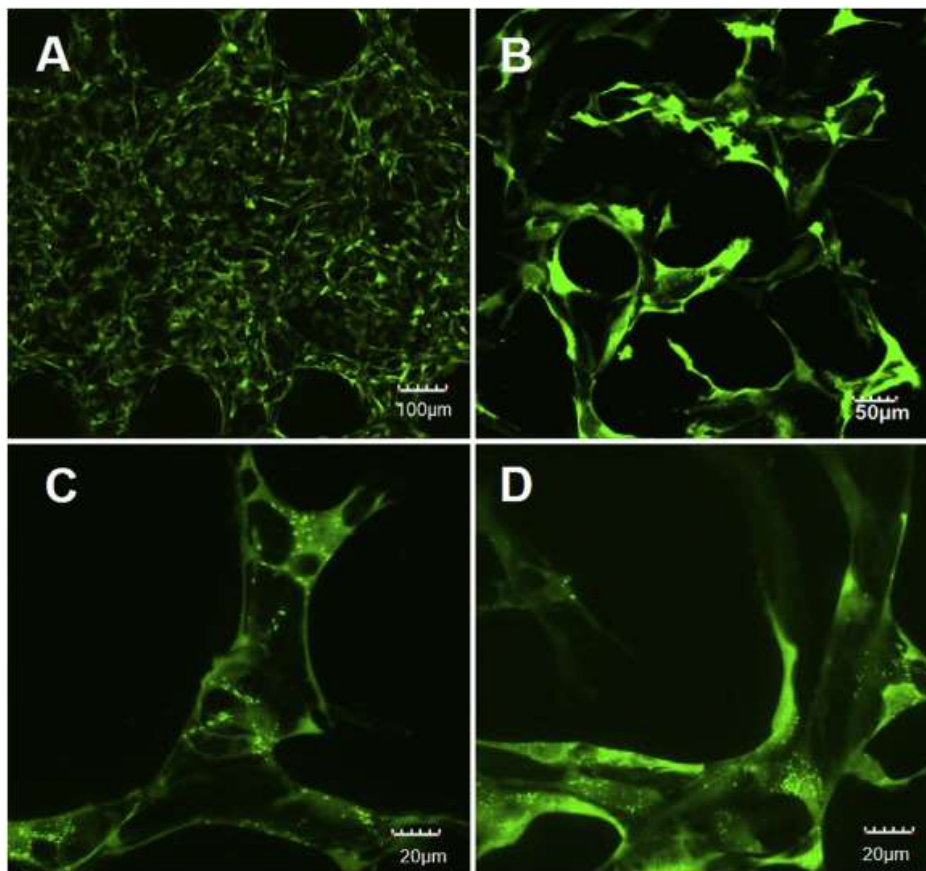


Image: A microscope observed the progress in the development of HBMECs seeded in a microfluidic chip. (A) Fluorescent HBMECs 24 h after being seeded (B) HBMECs 48 h after being seeded shows cell extensions and differentiation gradually; (C, D) the vascular lumen 3D structures were formed along with cell proliferation (Green Fluorescence-Green fluorescence protein). Data courtesy of Yang Wu from Wuhan Polytechnic University (<https://doi.org/10.1016/j.ecoenv.2019.110077>)

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Characterization of the cells

- Cytoplasmic VWF / Factor VIII: >95% positive by immunofluorescence
- Cytoplasmic uptake of Di-I-Ac-LDL: >95% positive by immunofluorescence
- Cytoplasmic PECAM1 >95% positive by immunofluorescence

HBMECs are negative for HIV-1, HBV, HCV, and mycoplasma.

Recommended Products

- ENDO-Growth Media – NB-26-00280-500ML
 - Contains 475 ml of ENDO-Basal Media and 25 ml of ENDO-Growth Supplement combined. Which is freshly prepared for your convenience
- OR
- ENDO-Growth Kit – NB-26-00100
 - Contains 475 ml of ENDO-Basal Media and 25 ml of ENDO-Growth Supplement in separately to be mixed to make growth media
- Smooth Coat Solution – NB-26-001598-20ML
 - Biocompatible complex of extracellular matrix binding solution
- OR
- AlphaBioCoat Solution – NB-26-01690
 - Premium Smooth Coat Solution. Biocompatible complex of extracellular matrix binding solution with growth factors. Ideal for culturing cells from frozen.
- Cell Detachment Solution – NB-26-01693
 - Contains protease and collagenase activities in an isotonic, phosphate buffer solution with EDTA to detach primary cells and cell lines
- 1X Phosphate Buffer Solution - NB-26-00723-100ML

Shipping

Shipped on dry ice frozen in a vial.

Handling of Arriving Cells

Store in liquid nitrogen to keep the cells frozen or thaw cells according to the protocol for culture.

Note: Handling human derived products is potentially biohazardous. Although each cell strain tests negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate, therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.

SUBCULTURE PROTOCOL

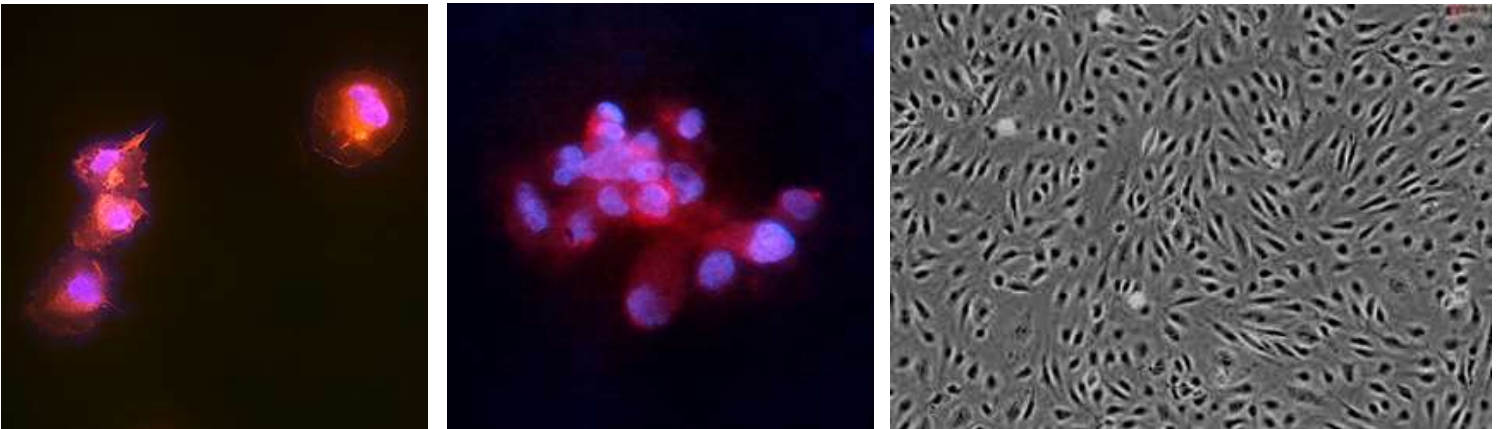
Note: If you have any questions or need clarification regarding the protocol for culturing these cells, please contact us at tech@neo-biotech.com

1. Coating T25 flasks:
 - a. Add 2 ml AlphaBioCoat Solution (NB-26-01690) into a T25 flask and ensure entire interior surface is coated with solution. After 30 minutes, dispose of Smooth Coat Solution by aspiration. Gently rinse and aspirate flask with phosphate buffer solution (NB-26-00723-100ML). The flask is now ready for use (no need for overnight incubation when coated with NB-26-01690)
 - b. If you are using the coated flask the same day, add about 4 ml of Endo-Growth media to the coated flask. *If the media changes color from pink to yellow, aspirate and discard the media. Add 4ml of fresh media to the coated flask.
2. Thaw the cells in a 37°C water bath. Once you see a small amount of ice left in the vial, spray the vial with 70% Ethanol and wipe it down.

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3. Transfer the vial into your Biosafety cabinet.
4. Using a 2 or 5ml pipet, pipet the cells out of the vial.
5. Transfer your cell suspension in to your coated flask (which contains 4 ml media).
6. You should have a total working volume of 5ml of cell suspension in the flask; close the cap. Make sure cells are evenly distributed in the flask by moving the flask left and right five times. Move it up and down for an additional five times.
7. Place flask in a 37°C incubator with 5% CO₂. If flask is not vented, please loosen cap.
8. Change media after 48 hours.
9. Place flask in 37°C incubator until cells are at 90% confluence. Change media every 2 days.
10. When flask is at 90% confluence, aspirate media from flask.
11. Rinse T25 flask containing cells with 5 ml 1XPBS (cat#PBS300).
12. Gently aspirate out the PBS after rinsing, and discard.
13. Add 2ml of RT trypsin/ EDTA or Cell Detachment Solution (ADF001) to T25 flask containing cells (ensure entire interior surface is covered).
14. Place T25 flask containing cells into 37°C incubator for 1 or 2 minutes (cells will normally come off of the surface within 1 or 2 minutes).
15. Suspend the cells with 15ml of ENDO-Growth medium (MED001) and transfer equally into 3 pre-coated T25 flasks (the cells are now at a subculture ratio of 1:3).
16. There is no need to spin cells during subculture.
17. Proliferating cell culture: ENDO-Growth medium (MED001) should be changed every 2 days. The cells normally become confluent within 7 days (when split at a 1:3 ratio)
18. Use ENDO- Basal media (MED002) containing 0.5% FBS to induce quiescent cells (after 18-24 hours).



Images: **Left:** HBMECs stained with CD146. **Middle:** HBMECs stained with Claudin-5. **Right:** Human Brain Microvascular Endothelial Cells in Culture.

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