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## Minute™ Cytoplasmic and Nuclear Extraction Kit for Cells

Catalog number: SC-003

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### Description

Invent Biotechnologies Minute™ Cytoplasmic and Nuclear Extraction Kit for Cells is composed of optimized cytoplasmic extraction buffer, nuclear protein extraction buffer and protein extraction filter cartridges with 2.0 ml collection tubes. The kit is designed to rapidly separate native cytosol and nuclear proteins from **mammalian cells and protoplasts of plant, bacteria, yeast and fungus**. Due to the use of the protein extraction filter cartridges separation of cytoplasmic and nuclear proteins can be accomplished in less than 15 min.

### Application

The kit is designed to rapidly extract native cytoplasmic and nuclear proteins from cultured cells for applications such as SDS-PAGE, immunoblotting, ELISA, IP, protein localization, gel mobility shift assays, 2-D gels and other applications. This kit provides the most rapid method currently available for fractionation of native cytoplasmic and nuclear proteins.

### Kit Components

1. 25 ml cytoplasmic extraction buffer
2. 25 ml nuclear extraction buffer
3. 50 protein extraction cartridges
4. 50 collection tubes with cap

**Shipping:** This kit is shipped at ambient temperature **Storage:** Store the kit at 4°C upon arrival.

### Important Product Information

The use of protease inhibitors is not necessary prior to extraction. However if downstream application takes significant amounts of time or the protein extract will be stored for longer period of time, addition of protease inhibitor to extracted lysate is recommended. The nuclear extraction buffer contains 300 mM salt, for some applications, dilution or desalting of the extract may be needed. For determination of protein concentration, BCA kit (Pierce) is recommended. To study protein phosphorylation, **phosphatase inhibitors** (such as PhosStop from Roche) should be added to lysis buffer prior to use.

### Additional Materials Required

1 X PBS  
Vortexer  
Table-Top Microcentrifuge  
BCA Protein Assay Kit (Pierce, Cat #: 23227)



## A. Cultured Cells in suspension (including protoplasts from plant, bacteria, yeast and fungus)

1. Harvest cells in suspension by low speed centrifugation (500 X g for 3 min). Wash the cell in cold PBS once.
2. Transfer the cells to a 1.5 ml microcentrifuge tube and pellet the cells by centrifugation at 500 X g for 1 min; aspirate the supernatant completely.
3. Add appropriate amounts of cytoplasmic extraction buffer to cell pellets (Table 1), vortex the tube vigorously for 15 seconds, incubate on ice for 5 min and vortex briefly (**NOTE: longer incubation may be needed if minimum contamination in nuclear fraction is more desirable**). Go to Cytoplasmic and Nuclear Protein Extraction Procedures below.

## B. Adherent cells

1. Grow adherent cells to 90-100% confluence and wash the cells twice in the tissue culture plates, dishes or flasks with cold PBS, aspirate the buffer completely.
2. Add appropriate amounts of cytoplasmic extraction buffer (Table 2), swirl to distribute the lysis buffer over the entire surface of tissue cultures, place the tissue culture on ice for 5 min (**NOTE: longer incubation may be needed if minimum contamination in nuclear fraction is more desirable**). Scrape the lysed cells with a pipette tip or with a transfer pipette and transfer cell lysate to pre-chilled 1.5 ml microcentrifuge tube. Vortex the tube vigorously for 15 seconds. Go to Cytoplasmic and Nuclear Protein Extraction Procedures below.

**Table 1. Buffer volume for different packed cell volume**

| Packed Cell Volume (µl) | Cytoplasmic Extraction Buffer (µl) | Nuclear Extraction Buffer (µl) |
|-------------------------|------------------------------------|--------------------------------|
| 5                       | 50                                 | 25                             |
| 10                      | 100                                | 50                             |
| 20                      | 200                                | 100                            |
| 50                      | 500                                | 250                            |

\*For NIH3T3 and 293T cells 10 µl packed cell volume is equivalent to 10<sup>6</sup> cells

**Table 2. Buffer Volume for Different Amount of Adherent Cells**

| Containers               | Cytoplasmic Extraction Buffer (µl) | Nuclear Extraction Buffer (µl) |
|--------------------------|------------------------------------|--------------------------------|
| 24-well plate            | 80                                 | 25                             |
| 6-well plate             | 300                                | 150                            |
| 25 cm <sup>2</sup> flask | 500                                | 250                            |

## Cytoplasmic and Nuclear Protein Extraction Procedures

1. Centrifuge the tube for 5 min at top speed in a microcentrifuge at 4°C.
2. Transfer the supernatant (cytosol fraction) to a fresh pre-chilled 1.5 ml tube (optional: wash the pellet with 0.5 ml cold PBS to reduce contamination of cytosolic proteins). Add appropriate amounts of



nuclear extraction buffer to the pellet, vortex vigorously for 15 seconds, incubate the tube on ice for one min. Repeat the 15 second vortexing and one min incubation 4 times.

3. Immediately transfer/pour the nuclear extract to a pre-chilled filter cartridge with collection tube and centrifuge at top speed (14,000-16,000 xg) in a microcentrifuge for 30 seconds. Discard the filter cartridge. Store nuclear extract at -80°C until use. Typical protein yield is about 1.5-2.5 mg/ml.

## Troubleshooting

| Problem   | Solution  |
|---|---|
| Low protein concentration   | Increase amounts of cells/tissues or decrease amount of cell lysis buffer |
| Low protein activity  | Keep lysate cold/add protease inhibitors                                  |
| Significant contamination of nuclear fraction by cytosolic proteins | Add NP-40 to cytosolic extraction buffer to a final concentration of 0.1% |