NeoBio Mount Universal

(Water Based, universal, coverslip free)

#Cat: NB-23-00157-1	**Size: 100ml liquid form.**
#Cat: NB-23-00157-2	**Size: 18ml liquid form.**

Intended Use

NeoBio Mount Universal is water-based mounting medium for immunohistology. It is used for the permanent preservation of the following chromogens or dyes. Simply apply drops of NeoBio Mount Universal on the tissue section a liquid polymer coverslip will form. It does not need coverslip. This universal mounting media maintains the original stain intensity and characteristics. It demonstrates excellent optical clarity.

Compatible with:

1. Horseradish peroxidase chromogens:
 - Diaminobenzidine (DAB)
 - 3-amino, 9-ethylcarbazole (AEC)
 - 4-chloro-1-naphthol

2. Alkaline phosphatase chromogens:
 - 5-bromo, 4-chloro,indolylphosphate/ iodonitrotetrazolium (BCIP/INT)
 - 5-bromo, 4-chloro, 3-indolylphosphate/nitro- bluetetrazolium (BCIP/NBT)
 - Naphthol AS-MX phosphate/Fast Red TR salt

3. Fluorescent specimen:
 - Fluorescein
 - Rhodamine
 - Texas Red
 - FluoroBlue

NOT Compatible with:

1. Fluorescent Specimen stained with:
 - Phycoerythrin
 - Phycocyanin
 - Allophycocyanin

2. Conterstains:
 - Nuclear Fast Red
 - BC-50
 - Light Green

3. Tetramethylbenzidine (TMB)
4. Eosin

Components

Contains less than 0.05% sodium azide.

Storage

Store at 2-8°C for longer shelf-life

Recommended Protocol

1. Recommended maximum section thickness is 5-8 microns. Thicker sections may produce air pockets
2. Must apply NeoBio Mount Universal when tissue is wet. Air bubbles may be formed when tissue is dry
3. Rinse slide with **DISTILLED OR DEIONIZED WATER**, touch the edges of slide on a paper towel to remove excess water and quickly wipe the back of the slide.
4. Invert dropper bottle to displace bubbles from the tip-end.
5. Squeeze out the first drop onto a paper towel to remove of air bubble on the tip.
6. Apply 3 drops of NeoBio Mount Universal to cover the tissue section. DO NOT coverslip on top of the NeoBio Mount Universal.
7. Rotate the slides to allow NeoBio Mount Universal spread evenly to cover the tissue section.
8. Place slides horizontally in an oven at 40-50°C for at least 30 minutes or leave it at room temperature until slides are thoroughly dried. Slow dry at room temperature will help to eliminate the air bubbles. When use with fluorescent specimen, bake at 60-70°C for 20 minutes.
9. Hardened NeoBio Mount Universal forms an impervious polymer barrier to organic solvent.
10. Allow sufficient time of drying slides before storage.
11. For maximum retention of fluorescence, slides should be kept in dark place at 4°C.
12. Do not use oil directly on the top of dried NeoBio Mount Universal. After saline rinse, shake off excess liquid from the slide and quickly wipe the back of the slide.
2. Invert dropper bottle to displace bubbles from the tip-end.
3. Squeeze out the first drop onto a paper towel to remove of air bubble on the tip.
4. Add a few drops of GB-Mount on the tissue section on the slide.
5. Carefully lay down coverglass to cover the tissue section.
6. Gently press out any noticeable air bubbles. If there are many bubbles on the slide, one may soak the slide in saline, remove the coverslip, and repeat the coverslipping procedure.
7. The slide may be viewed immediately after coverslipping, be careful not to dislodge the coverslip before the slide has dried.
8. Allow sufficient time of drying slides before storage.
9. For maximum retention of fluorescence, slides should be kept in dark place at 4°C.

Removal of Coverslip: If the coverslip has to be removed, soak the slide in warm (37°C) water for 5-10 minutes until the mounting medium is dissolved. Rinse slide with warm water to remove all mounting medium. The slide can be remounted again.

For Research Use Only. Not for Diagnostic or Therapeutic Use.

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